C10N	0.3204 (3)	-0.3140 (3)	1.332(1)	0.049 (2)
C11	0.4966 (3)	0.1870 (3)	1.0184 (9)	0.045 (2)
C12	0.4813 (3)	0.2607 (3)	1.0988 (10)	0.050 (2)
C13	0.4620 (3)	0.3081 (3)	0.9151 (9)	0.040 (2)
C14	0.3994 (2)	0.2767 (3)	0.7914 (9)	0.034 (2)
C15	0.3741 (3)	0.3365 (3)	0.6497 (9)	0.047 (2)
C16	0.3811 (3)	0.4012 (3)	0.795 (1)	0.058 (2)
C17	0.4332 (4)	0.3786 (3)	0.973 (1)	0.054 (2)
C18	0.5166 (3)	0.0767 (3)	0.660(1)	0.055 (2)
C19	0.5275 (3)	0.3232 (3)	0.776 (1)	0.060 (2)

Table 4. Selected	geometric parameters	(À	Å)	for	BENA
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O1-C3	1.234 (7)	C2—C3	1.459 (9)
02-C17	1.201 (8)	C3-C4	1,464 (9)
O1N-C2N	1.333 (8)	C4C5	1.328 (8)
C1-C2	1.324 (8)	C5C6	1.501 (8)
C1-C10	1.511 (8)		

For both compounds, data collection: MSC/AFC Diffractometer Control Software (Molecular Structure Corporation, 1988); cell refinement: MSC/AFC Diffractometer Control Software; data reduction: TEXSAN PROCESS (Molecular Structure Corporation, 1992). Program(s) used to solve structures: SHELXS86 (Sheldrick, 1990) for ALNA; SIR88 (Burla et al., 1989) for BENA. For both compounds, program(s) used to refine structures: TEXSAN LS; software used to prepare material for publication: TEXSAN FINISH.

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Lists of structure factors, anisotropic displacement parameters, H-atom coordinates, bond distances and angles involving non-H atoms for ALNA and complete geometry for BENA have been deposited with the IUCr (Reference: KA1098). Copies may be obtained through The Managing Editor, International Union of Crystallography, 5 Abbey Square, Chester CH1 2HU, England.

References

- Burla, M. C., Camalli, M., Cascarano, G., Giacovazzo, C., Polidori, G., Spagna, R. & Viterbo, D. (1989). J. Appl. Cryst. 22, 389–393.
- Dryden, H. L., Webber G. M. & Wieczorek J. J. (1964). J. Am. Chem. Soc. 86, 742–743.
- Fried, J., Thoma, R. W. & Klingsberg, A. (1953). J. Am. Chem. Soc. 75, 5764–5765.
- Kálmán, A., Párkányi, L. & Argay, Gy. (1993). Acta Cryst. B49, 1039–1049.
- Marsheck, W. J., Kraychy, S. & Muir, R. D. (1972). Appl. Microbiol. 23, 72–77.
- Molecular Structure Corporation (1988). MSC/AFC Diffractometer Control Software. MSC, 3200 Research Forest Drive, The Woodlands, TX 77381, USA.
- Molecular Structure Corporation (1992). TEXSAN. TEXRAY Structure Analysis Package. MSC, 3200 Research Forest Drive, The Woodlands, TX 77381, USA.
- Nagasawa, M., Bae, M., Tamura, G. & Arima, K. (1969). Agric. Biol. Chem. 33, 1644–1650.
- Pauncz, J., Wix, G., Rados, M. & Alföldi, L. (1961). Hungarian Patent No. 148 094.
- Sheldrick, G. M. (1990). Acta Cryst. A46, 467-473.
- Wix, G., Büki, K. G., Tömörkény, E. & Ambrus, G. (1968). Steroids, 11, 401-413.

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DL-4-Hydroxy-3-methoxymandelic Acid

NOBUO OKABE, TAMAMI SUGA AND YOSHIKO KOHYAMA

Faculty of Pharmaceutical Sciences, Kinki University, Kowakae 3-4-1, Higashi Osaka, Osaka 577, Japan

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Abstract

In the title compound, DL-2-(4-hydroxy-3-methoxyphenyl)-2-hydroxyacetic acid, $C_9H_{10}O_5$, the acetic acid side chain adopts a roughly perpendicular orientation with respect to the phenyl ring. The molecules are linked together through hydrogen bonds of the type O—H···O.

Comment

The crystal structures of the catecholamines dopamine (Bergin & Carlström, 1968; Giesecke, 1980), adrenaline (Andersen, 1975a), noradrenaline (Andersen, 1975b; Carlström & Bergin, 1967) and their analogues (Barlow, Johnson, Howard, Walton & Koellner, 1989; Seiler, Markstein, Walkinshaw & Boelsterli, 1989) have been determined. It is also important to clarify the detailed structure of catecholamine metabolites in order to study catecholamine action as well as metabolism. In this respect the structures of the dopamine metabolites 3-methoxytyramine (Okabe, Mori & Sasaki, 1991; Okabe & Mori, 1992) and homovanillic acid (Okabe, Hatanaka & Sasaki, 1991), and the noradrenaline metabolite normetanephrine (Pattanayek, Dattagupta, Bhattacharyya & Saha, 1984) have been reported. We report here the crystal structure of the title compound, (I), which is the principal metabolite of adrenaline and noradrenaline (Grodsky, 1983).



The acetic acid side chain is oriented roughly perpendicularly to the phenyl ring [torsion angle C(2)—C(1)—C(7)—C(8) $-63.8(2)^{\circ}$]. This conformational feature of the molecule resembles that observed for catecholamines and the corresponding amines (Barlow, Johnson, Howard, Walton & Koellner, 1989) as well as the catecholamine metabolites normetanephrine (Pattanayek, Dattagupta, Bhattacharyya & Saha, 1984), homovanillic acid (Okabe, Hatanaka & Sasaki, 1991) and 3-methoxytyramine (Okabe & Mori, 1992). Two hydroxyl groups and the carboxyl group participate

in hydrogen bonding: $O(7) \cdots O(4)(\frac{1}{2} - x, \frac{1}{2} + y, \frac{3}{2} - z)$ 2.761 (2), $O(4) \cdots O(82)(1 - x, -y, 2 - z)$ 2.931 (2) and $O(81) \cdots O(7)(\frac{1}{2} - x, -\frac{1}{2} + y, \frac{5}{2} - z)$ 2.718 (2) Å.



Fig. 1. Perspective view of the title compound with the atomic numbering scheme. Displacement ellipsoids are shown at the 50% probability level and H atoms are drawn as spheres of arbitrary size. The molecule is viewed down the b axis.

Experimental

Crystals were obtained by evaporation from 70% ethanol. The density D_m was measured by flotation in CCL₄/C₆H₆.

Crystal data

Mo $K\alpha$ radiation C₉H₁₀O₅ $M_r = 198.18$ $\lambda = 0.71069 \text{ Å}$ Monoclinic Cell parameters from 25 reflections $P2_1/n$ $\theta = 15.25 - 19.30^{\circ}$ a = 10.086(3) Å $\mu = 0.112 \text{ mm}^{-1}$ b = 8.829(2) Å T = 296 Kc = 10.503 (2) ÅPrism $\beta = 103.43 (2)^{\circ}$ $0.30 \times 0.30 \times 0.20$ mm $V = 909.6 (4) \text{ Å}^3$ Colorless Z = 4 $D_x = 1.447 \text{ Mg m}^{-3}$ $D_m = 1.439 (3) \text{ Mg m}^{-3}$ Data collection

 $\theta_{\rm max} = 27.5^{\circ}$

 $h = 0 \rightarrow 13$

 $k = 0 \rightarrow 11$

 $l = -13 \rightarrow 13$

3 standard reflections

monitored every 150 reflections

intensity decay: 0.40%

frequency: 60 min

Rigaku AFC-5R diffractometer $\omega/2\theta$ scans Absorption correction: none 2349 measured reflections 2229 independent reflections 1553 observed reflections $[I > 1.5\sigma(I)]$ $R_{int} = 0.021$

Refinement

Refinement on F	$(\Delta/\sigma)_{\rm max} = 0.002$
R = 0.047	$\Delta \rho_{\rm max} = 0.22 \ {\rm e} \ {\rm \AA}^{-3}$
wR = 0.054	$\Delta \rho_{\rm min} = -0.29 \ {\rm e} \ {\rm \AA}^{-3}$
S = 1.71	Extinction correction: none
1553 reflections	Atomic scattering factors
127 parameters	from International Tables
H-atom parameters not	for X-ray Crystallography
refined	(1974, Vol. IV)
$w = 4F_o^2/\sigma^2(F_o^2)$	

Table 1. Fractional atomic coordinates and equivalent isotropic displacement parameters (Å²)

Bea =	$(8\pi^2/3)\sum_i\sum_iU_{ii}a_i^*a_i^*\mathbf{a}_i\cdot\mathbf{a}_i.$	
- eq	$(on for) = f = f o f a_1 a_2 = f a_2$	

	x	y	Z	Bea
O(3)	0.5745 (1)	0.1564 (2)	0.8562(1)	3.48 (6)
O(4)	0.4195 (2)	0.0196 (2)	0.6483 (1)	3.98 (7)
0(7)	0.2028 (2)	0.3758 (2)	1.0824 (1)	3.62 (6)
O(81)	0.1685(1)	0.0097 (2)	1.1893 (1)	3.62 (7)
O(82)	0.3408 (2)	0.1719 (2)	1.2553 (1)	3.45 (6)
C(1)	0.2410 (2)	0.1644 (2)	0.9448 (2)	2.55 (7)
C(2)	0.3802 (2)	0.1871 (2)	0.9560 (2)	2.61 (7)
C(3)	0.4401 (2)	0.1391 (2)	0.8576 (2)	2.61 (7)
C(4)	0.3617 (2)	0.0654 (2)	0.7485 (2)	2.81 (8)
C(5)	0.2251 (2)	0.0407 (3)	0.7385 (2)	3.11 (8)
C(6)	0.1646 (2)	0.0920 (3)	0.8366 (2)	3.07 (8)
C(7)	0.1760 (2)	0.2198 (2)	1.0537 (2)	2.75 (8)
C(8)	0.2383 (2)	0.1336 (2)	1.1786 (2)	2.67 (8)
C(9)	0.6599 (2)	0.2301 (3)	0.9668 (2)	3.7 (1)

Table 2. Selected geometric parameters (Å, °)

O(3)—C(3)	1.368 (2)	C(1)—C(2)	1.397 (3)
O(3)-C(9)	1.432 (2)	C(1)—C(6)	1.374 (3)
O(4)—C(4)	1.377 (2)	C(1)—C(7)	1.524 (3)
O(7) -C(7)	1.422 (3)	C(2)—C(3)	1.379 (3)
O(81)—C(8)	1.320 (2)	C(3)—C(4)	1.393 (3)
O(82)C(8)	1.203 (2)	C(4)—C(5)	1.375 (3)
C(7)—C(8)	1.521 (3)	C(5)C(6)	1.389 (3)
C(3)	116.9 (2)	C(3)—C(4)—C(5)	120.3 (2)
C(2) - C(1) - C(6)	119.7 (2)	C(4)—C(5)—C(6)	119.8 (2)
C(2) - C(1) - C(7)	119.3 (2)	C(1)C(6)C(5)	120.5 (2)
C(6) - C(1) - C(7)	121.0 (2)	O(7)—C(7)—C(1)	112.1 (2)
C(1) - C(2) - C(3)	120.0 (2)	O(7)C(7)C(8)	106.4 (2)
O(3)-C(3)-C(2)	125.5 (2)	C(1)—C(7)—C(8)	108.8 (2)
O(3) - C(3) - C(4)	114.7 (2)	O(81)-C(8)-O(82)	124.7 (2)
C(2) - C(3) - C(4)	119.8 (2)	O(81)—C(8)—C(7)	111.5 (2)
O(4)-C(4)-C(3)	120.4 (2)	O(82)C(8)C(7)	123.8 (2)
O(4)-C(4)-C(5)	119.3 (2)		

Data collection and cell refinement: Rigaku MSC/AFC Data Collection and Refinement Software (Rigaku Corporation, 1988). Data reduction: TEXSAN (Molecular Structure Corporation, 1985). Structure solution: SHELXS86 (Sheldrick, 1985) and DIRDIF (Beurskens, 1984). Structure refinement and molecular graphics: TEXSAN.

Lists of structure factors, anisotropic displacement parameters, Hatom coordinates and complete geometry, including torsion angles, have been deposited with the IUCr (Reference: AS1143). Copies may be obtained through The Managing Editor, International Union of Crystallography, 5 Abbey Square, Chester CH1 2HU, England.

References

Andersen, A. M. (1975a). Acta Chem. Scand. Ser. B, 29, 239-244. Andersen, A. M. (1975b). Acta Chem. Scand. Ser. B, 29, 871-876.

- Barlow, R. B., Johnson, O., Howard, J. A. K., Walton, D. C. & Koellner, G. (1989). Acta Cryst. B45, 396-404.
- Bergin, R. & Carlström, D. (1968). Acta Cryst. B24, 1506-1510.
- Beurskens, P. T. (1984). DIRDIF. Direct Methods for Difference Structures – an Automatic Procedure for Phase Extension and Refinement of Difference Stucture Factors. Technical Report 1984/1. Crystallography Laboratory, Toernooiveld, 6525 ED Nijmegen, The Netherlands.
- Carlström, D. & Bergin, R. (1967). Acta Cryst. 23, 313-319.
- Giesecke, J. (1980). Acta Cryst. B36, 178-181.
- Grodsky, G. M. (1983). Harper's Review of Biochemistry, 19th ed., edited by D. W. Martin, P. A. Mayes & V. W. Rodwell, pp. 494– 497. Singapore: Maruzen Asian.
- Molecular Structure Corporation (1985). TEXSAN. TEXRAY Structure Analysis Package. MSC, 3200 Research Forest Drive, The Woodlands, TX 77381, USA.
- Okabe, N., Hatanaka, Y. & Sasaki, Y. (1991). Acta Cryst. C47, 2181-2183.
- Okabe, N. & Mori, S. (1992). Acta Cryst. C48, 1698-1699.
- Okabe, N., Mori, S. & Sasaki, Y. (1991). Acta Cryst. C47, 1448–1450. Pattanayek, R. R., Dattagupta, J. K., Bhattacharyya, S. C. & Saha, N. N. (1984). Acta Cryst. C40, 294–297.
- Rigaku Corporation (1988). Rigaku MSC/AFC Data Collection and Refinement Software. Rigaku Corporation, Tokyo, Japan.
- Seiler, M. P., Markstein, R., Walkinshaw, M. D. & Boelsterli, J. J. (1989). Mol. Pharmacol. 35, 643–651.
- Sheldrick, G. M. (1985). SHELXS86. Program for the Solution of Crystal Structures. Univ. of Göttingen, Germany.

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3-(p-Hydroxyphenyl)propionic Acid

NOBUO OKABE AND TAMAMI SUGA

Faculty of Pharmaceutical Sciences, Kinki University, Kowakae 3-4-1, Higashiosaka, Osaka 577, Japan

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Abstract

The title compound, $C_9H_{10}O_3$, has a fully extended propionic side chain in a *trans* configuration; the plane of the side chain is almost perpendicular to the phenylring plane. The molecules are held together by two kinds of hydrogen bonds between hydroxyl groups and between carboxyl groups.

Comment

p-Hydroxyphenylpropionic acid, (I), is well known as one of the intermediates of tyrosine metabolites such as p-hydroxyphenyl pyruvic acid, p-hydroxyphenyllactic acid, p-hydroxylphenylacrylic acid or p-hydroxyphenylacetic acid. The excretion of p-hydroxyphenylpropionic acid increases in patients with gastrointestinal diseases such as cystic fibrosis, coeliac disease or intestinal resection (van der Heiden, Wauters, Ketting, Duran & Wadman, 1971). On the other hand, *p*-hydroxyphenylpropionic acid inhibits peptic hydrolysis by pepsin (Schlamowitz, Shaw & Jackson, 1968). It also binds strongly to peroxidases which catalyse the oxidation of a large number of organic substances (Casella *et al.*, 1991). The present study was performed to find basic conformational features of the title compound for further investigation of its physiological function.



The molecule has a fully extended propionic side chain in a *trans* configuration $[C(1)-C(7)-C(8)-C(9) = 177.8 (2)^{\circ}]$. The plane of the side chain is almost perpendicular to the phenyl plane $[C(6)-C(1)-C(7)-C(8) = 112.6 (2)^{\circ}]$. Molecules are held together by two kinds of O-H···O intermolecular hydrogen bonds between two hydroxyl groups and between two carboxyl groups: O(4)-H(4)···O(4)(1 - x, 1 - y, 2 - z) 2.927 (3); O(91)-H(91)···O(92)(2 - x, 1 - y, 1 - z) 2.662 (2) Å.



Fig. 1. Perspective view of the title compound with the atomic numbering. Ellipsoids for non-H atoms correspond to 50% probability.

Experimental

Crystal data $C_9H_{10}O_3$ Mo $K\alpha$ radiation $M_r = 166.18$ $\lambda = 0.71069 \text{ Å}$ Monoclinic Cell parameters from 25 $P2_1/c$ reflections $\theta = 22.6 - 24.75^{\circ}$ a = 11.356(2) Å b = 5.358(1) Å $\mu = 0.094 \text{ mm}^{-1}$ T = 296 Kc = 14.122(2) Å $\beta = 105.94^{\circ}$ Needle $V = 826.3 (3) \text{ Å}^3$ $0.40 \times 0.20 \times 0.10$ mm Z = 4Colourless $D_x = 1.336 \text{ Mg m}^{-3}$ Crystal source: evaporation from water

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